

Induction of host resistance with plant defense activators against white rust [*Albugo candida*] of Indian mustard

Hanuman Singh^{*1}, RS Ratnoo²

¹College of Horticulture and Forestry, Jhalawar, Agriculture University, Kota 324001 Rajasthan, India

²Maharana Pratap University of Agriculture and Technology, Udaipur 313001 Rajasthan, India

*Corresponding author: rathore.hanuman99@gmail.com

(Received: 19 April 2021, Revised: 8 June 2021; Accepted: 10 June 2021)

Abstract

In plants systemic resistant can be induced through biotic or abiotic plant defense inducers. Efficacy of biotic and abiotic plant activators was tested against *Albugo candida* causing white rust on Indian mustard cultivar RH 406 under artificial epiphytotic conditions during two consecutive cropping seasons. Maximum average size of pustules were recorded in check (3.92 mm) followed by zinc sulphate (3.27 mm), *Trichoderma viride* (3.14 mm). Among the abiotic agents, maximum reduction in the size of the pustules were recorded in salicylic acid (47.01 %) followed by calcium sulphate (41.28 %) and potassium chloride (40.06 %). Overall number of pustules were recorded maximum in check (5.78), followed by zinc sulphate (5.39) and *T.viride* (5.16). Among the bio agents, *Pseudomonas fluorescens* reduced number of pustules by 25.96% over the check. Among the abiotic agents, salicylic acid recoded superior in all treatments over the control which reduced number of pustules 38.65% over the check followed by calcium sulphate (32.50%). Among the abiotic agents, salicylic acid reduce the percent disease index by 30.11% over the check followed by calcium sulphate (22.54%) and minimum percent disease index reduction was recorded in zinc sulphate was 5.93%.

Keywords: *Albugo candida*, Indian mustard, resistance, white rust

Introduction

Plants exhibit numerous responses when challenged by pathogens. Some of these involve the activation of host defense genes that bring about physical and biochemical changes in the host (Jones and Dangle, 2006). It is well established that resistance can be induced in plants by biotic as well as by abiotic agents (Kessman *et al.*, 1994; Kombrink and Sommsich, 1995; Sticher *et al.*, 1997). In recent years, a new group of compounds that activate host defense mechanism and protect the plant against pathogens has been developed to manage crop diseases. These chemicals are identified as “plant defense activators” or “plant activators” (Romero *et al.*, 2001). Host plant defense can be induced by the application of non-pathogenic microorganisms (Vishwanath *et al.*, 1999; Singh *et al.*, 1999) and certain abiotic activators such as salicylic acid (Spletzer and Enyedi, 1999). Most commonly used chemicals inducers is salicylic acid, which appears to mimic the systemic effects of localized infection in plant system (Safari *et al.*, 2013). External application of salicylic acid can induce systemic acquired resistance (Hammerschmidt, 1999). One of the potential management methods is the use of systemic acquired resistance to trigger host defense mechanisms, which would not involve the application of hazardous compounds to plants (Durrant and Dong, 2004).

Induced resistance has been exploited widely for the management of plant diseases (Gorlach *et al.*, 1996). Salicylic acid reported effective inducers of plant defense in the host plant system (Yalpani and Raskin, 1993). In addition, numerous microorganisms applied to the leaves or roots of plants may induce systemic or local resistance (Liu *et al.*, 1995). Such resistance is reported to be active against many types of organisms (Matheron and Porchas, 2002).

Biological plant defense inducers provide systemic resistance to plants infested by various fungal and bacterial phytopathogens. Biological plant defense inducers such as *Trichoderma viride*, *Pseudomonas*, *Bacillus*, *Serratia*, nonpathogenic strains of *Fusarium* and yeast have been developed as commercial product to combat various plant diseases (Droly *et al.*, 2002; Benhamon and Garand, 2001; Verhagen *et al.*, 2004; Howell *et al.*, 2000). Biocontrol activity of *Trichoderma* based biocontrol agents inheres in their ability to orchestrate several biochemical pathways in diseased plants (Surekha *et al.*, 2014).

There are various options available for the farmers to protect their crop from the disease. Some options include development of resistant cultivars, biological control, crop rotation, tillage, and chemical pesticides. Disease control

is largely based on the use of fungicides, bactericides, and insecticides. These chemical compounds toxic to plant invaders, causative agents, or vectors of plant diseases. However, the hazardous effect of these chemicals or their degradation products on the environment and human health strongly necessitates the search for new, harmless means of disease control. There must be some natural phenomenon of induced resistance to protect plants from disease. Elicitors are compounds, which activate chemical defense in plants. Various biosynthetic pathways are activated in treated plants depending on the compound used. Commonly tested chemical elicitors are salicylic acid, methyl salicylate, benzothiadiazole, benzoic acid, chitosan, and so forth

which affect production of phenolic compounds and activation of various defense-related enzymes in plants (Thakur and Sohal, 2013). The present study was undertaken to explore the possibility of utilizing induced host resistance as a realistic alternative to classical fungicides in white rust management.

Materials and Methods

First spray plant defense activators in the mustard variety RH-406 when plants shows first symptom of the disease in each treatment with their respective concentration using a randomized block design. One standard chemical check, Metalaxyl @ 0.10, 0.20 and 0.30 per cent and one sterile distilled water check was also maintained in three replications.

Table 1: Biotic and abiotic activators and their concentration under field study

Treatments	Biotic and abiotic agents	Concentration (%)
T ₁	<i>Trichoderma viride</i>	1.00
T ₂	<i>Pseudomonas fluorescens</i>	1.00
T ₃	Salicylic acid	0.25
T ₄	Borax (Na ₂ B ₄ O ₇ ·10H ₂ O)	0.50
T ₅	Potassium sulphate (K ₂ SO ₄)	1.00
T ₆	Calcium sulphate (CaSO ₄)	1.00
T ₇	Metalaxyl	0.10
T ₈	Metalaxyl	0.20
T ₉	Metalaxyl	0.30
T ₁₀	Potassium chloride (KCl)	1.00
T ₁₁	Zinc sulphate (ZnSO ₄ ·7H ₂ O)	0.50
T ₁₂	Check	-

Observations recorded

Size of pustule on leaves

Diameter of randomly selected five leaves was measured in millimeter with the help of plastic scale and average size of pustule was calculated and recorded at ten days interval.

Number of pustules

Numbers of pustules were counted as pustules per 25 mm² leaf area of randomly selected five leaves of plant. The observations were recorded on five leaves and average number of pustules was then calculated per 25 mm² leaf area.

Per cent disease index on leaf

The per cent disease index on leaf due to white rust was recorded at 10 days interval up to 90 days after sowing (DAS) by using of 0-5 rating scale given by Biswas *et al.*, 2011; Tirmali and Kolte (2012).

Ratings were given as per above mentioned rating scale

Numerical rating	Leaf area covered by the pustules (%)
0	No symptoms
1	1-10
2	11-25
3	26-50
4	51-75
5	>75

and white rust per cent disease index was calculated by using formula given by Wheeler (1969) and Mathur *et al.*, 2013. Observations were recorded by randomly selecting twenty five leaves from each replication and were rated as per the above rating scale and per cent disease index was calculated and statistically analyzed as described by Panse and Sukhatme (1985) for analysis of variance of randomized block design in order to test the significance of experimental results.

$$\text{White rust index (\%)} = \frac{\text{Sum of all numerical ratings}}{\text{No. of leaves examined} \times \text{max. grade of scale}} \times 100$$

Table 2: Effect of bio-stimulators and non-conventional chemicals on size of pustule on mustard cv. RH-406 under field conditions

Treatment	Concentration		Size of pustules (mm)										Overall		Reduction over check (%)
	(%)	2015-16					2016-17					Mean			
		70 DAS	80 DAS	90 DAS	Mean		70 DAS	80 DAS	90 DAS	Mean					
<i>Trichoderma viride</i>	1.00	0.89	3.01	5.49	3.13	0.91	3.03	5.50	3.15	3.14	19.96				
<i>Pseudomonas fluorescens</i>	1.00	0.56	2.21	4.65	2.47	0.59	2.23	4.67	2.50	2.49	36.59				
Salicylic Acid	0.25	0.41	1.81	3.97	2.06	0.44	1.83	3.99	2.09	2.08	47.01				
Borax	0.50	0.78	2.57	5.13	2.83	0.81	2.60	5.15	2.86	2.84	27.47				
K ₂ SO ₄	1.00	0.63	2.47	4.95	2.68	0.65	2.49	4.97	2.71	2.69	31.24				
CaSO ₄	1.00	0.47	2.06	4.33	2.29	0.50	2.09	4.35	2.31	2.30	41.28				
Metalaxy10.1%	0.10	0.33	1.60	3.33	1.76	0.36	1.63	3.35	1.78	1.77	54.89				
Metalaxy10.2%	0.20	0.31	1.53	3.21	1.68	0.34	1.55	3.23	1.71	1.69	56.76				
Metalaxy10.3%	0.30	0.31	1.46	3.12	1.63	0.33	1.49	3.13	1.65	1.64	58.15				
KCl	1.00	0.50	2.10	4.41	2.34	0.52	2.13	4.43	2.36	2.35	40.06				
ZnSO ₄	0.50	0.94	3.09	5.74	3.26	0.96	3.11	5.76	3.28	3.27	16.64				
Check	-	1.35	3.99	6.39	3.91	1.36	4.01	6.41	3.93	3.92	0.00				
SE±	-	0.01	0.06	0.11	-	0.02	0.03	0.04	-	-	-				
CD 5%	-	0.04	0.16	0.31	-	0.06	0.08	0.11	-	-	-				

DAS = Days after sowing

Table 3: Effect of bio-stimulators and non-conventional chemicals on number of white rust pustule on mustard cv. RH-406 under field conditions

Treatment	Concentration (%)	Number of pustules/25 mm ² area									
		2015-16					2016-17				
		70DAS	80DAS	90DAS	Mean	70DAS	80DAS	90DAS	Mean	Over all Mean	Reduction over check (%)
<i>Trichoderma viride</i>	1.00	3.47	5.07	6.93	5.16	3.53	5.20	6.73	5.16	5.16	10.77
<i>Pseudomonas fluorescens</i>	1.00	2.73	4.13	5.87	4.24	2.80	4.20	5.93	4.31	4.28	25.96
Salicylic Acid	0.25	2.33	3.33	4.87	3.51	2.40	3.40	4.93	3.58	3.54	38.65
Borax	0.50	3.13	4.80	6.53	4.82	3.20	4.87	6.60	4.89	4.86	15.96
K ₂ SO ₄	1.00	3.07	4.60	6.13	4.60	3.13	4.67	6.20	4.67	4.63	19.81
CaSO ₄	1.00	2.60	3.73	5.27	3.87	2.67	3.80	5.33	3.93	3.90	32.50
Metalaxyl 0.1%	0.10	1.87	3.07	4.27	3.07	2.00	3.13	4.33	3.16	3.11	46.15
Metalaxyl 0.2%	0.20	1.73	2.93	4.20	2.96	1.80	3.00	4.27	3.02	2.99	48.27
Metalaxyl 0.3%	0.30	1.73	2.80	4.20	2.91	1.80	2.87	4.27	2.98	2.94	49.04
KCl	1.00	2.87	3.93	5.40	4.07	2.93	4.00	5.47	4.13	4.10	29.04
ZnSO ₄	0.50	3.80	5.20	7.07	5.36	3.87	5.27	7.13	5.42	5.39	6.73
Check	-	4.07	5.80	7.40	5.76	4.13	5.87	7.40	5.80	5.78	0.00
SE _{me}	-	0.08	0.11	0.12	-	0.05	0.05	0.10	-	-	-
CD 5%	-	0.24	0.33	0.36	-	0.14	0.14	0.30	-	-	-

DAS = Days after sowing

Table 4: Effect of bio-stimulators and non-conventional chemicals on disease index of white rust on mustard cv. RH-406 under field conditions

Treatment	Concentration(%)	Disease index (%)										Over all Mean	Reduction over check (%)
		2015-16					2016-17						
		70 DAS	80 DAS	90 DAS	Mean	70 DAS	80 DAS	90 DAS	Mean				
<i>Trichoderma viride</i>	1.00	24.8(29.9)	42.1(40.5)	75.7(60.5)	47.6(43.6)	26.9(31.3)	43.5(41.3)	77.1(61.4)	49.2(44.5)	48.4	0.7		
<i>Pseudomonas fluorescens</i>	1.00	24.5(29.7)	41.9(40.3)	75.5(60.3)	47.3(43.5)	26.7(31.1)	43.2(41.1)	76.8(61.2)	48.9(44.4)	48.2	1.3		
Salicylic Acid	0.25	14.4(22.3)	27.2(31.4)	62.7(52.3)	34.8(36.1)	14.1(22.1)	26.7(31.1)	59.2(50.3)	33.3(35.3)	34.0	30.1		
Borax	0.50	17.6(24.8)	34.7(36.1)	68.0(55.6)	40.1(39.3)	20.0(26.6)	33.6(35.4)	66.7(54.7)	40.1(39.3)	40.1	17.7		
K ₂ SO ₄	1.00	18.4(25.4)	36.5(37.2)	69.6(56.5)	41.5(40.1)	21.6(27.7)	34.9(36.2)	70.1(56.9)	42.2(40.5)	41.9	14.1		
CaSO ₄	1.00	16.5(24.0)	32.3(34.6)	66.7(54.7)	38.5(38.3)	18.7(25.6)	29.6(33.0)	62.7(52.3)	37.0(37.5)	37.7	22.5		
Metalaxyl 0.1%	0.10	9.6(18.1)	24.5(29.7)	56.0(48.5)	30.0(33.2)	9.6(18.1)	24.8(29.9)	51.5(45.8)	28.6(32.3)	29.3	39.8		
Metalaxyl 0.2%	0.20	9.3(17.8)	24.3(29.5)	55.7(48.3)	29.8(33.1)	9.3(17.8)	24.5(29.69)	51.2(45.7)	28.4(32.2)	29.1	40.3		
Metalaxyl 0.3%	0.30	9.1(17.5)	24.0(29.3)	55.5(48.1)	29.5(32.9)	9.1(17.5)	24.3(29.51)	50.9(45.5)	28.1(32.0)	28.8	40.9		
KCl	1.00	20.5(27.0)	38.1(38.1)	72.0(58.1)	43.6(41.3)	22.4(28.3)	37.1(37.50)	72.5(58.4)	44.0(41.6)	43.8	10.1		
ZnSO ₄	0.50	22.1(28.1)	40.5(39.5)	72.8(58.6)	45.2(42.2)	23.7(29.2)	40.0(39.23)	75.7(60.5)	46.5(43.0)	45.8	5.9		
Check	-	25.6(30.4)	42.4(40.6)	76.0(60.7)	48.0(43.9)	27.2(31.4)	43.7(41.40)	77.3(61.6)	49.4(44.7)	48.7	0.0		
SSEm±	-	0.42	0.66	0.79	-	0.91	1.30	1.00	-	-	-		
CD 5%	-	1.21	1.90	2.26	-	2.61	3.74	2.88	-	-	-		

Values in parenthesis are angular transformed values; DAS= Days after sowing

Results and Discussion

The size of the pustules were enlarged from 70 days after sowing to 90 days after sowing in cv. RH-406 during 2015-16 and 2016-17 (Table 2). The maximum average size of pustules were recorded in check (3.92 mm) followed by zinc sulphate (3.27 mm), *Trichoderma viride* (3.14 mm). The maximum reduction of the size of the pustules were found in Metalaxyl 0.3% followed by Metalaxyl 0.2% was 58.15 mm and 56.76 mm, respectively. However different concentration of Metalaxyl was non-significant with each other at 70, 80 and 90 days after sowing during *Rabi* season 2015-16. Among the abiotic agents, maximum reduction in the size of the pustules were recorded in salicylic acid (47.01 %) followed by calcium sulphate (41.28 %) and potassium chloride (40.06 %). Among the bio agents which reduce the size of the pustules significantly over the check.

Minimum number of pustule were observed in Metalaxyl 0.2% and Metalaxyl 0.3% was 1.73 and 1.73, respectively (Table 3). However, the different concentrations of Metalaxyl were found at par with each other at 70, 80 and 90 days after sowing in 2015-16 and at 90 days after sowing in 2016-17. The reduction of the disease over the check in Metalaxyl 0.1%, 0.2% and 0.3% was found lowest were 46.15%, 48.27% and 49.04%, respectively. Overall number of pustules were recorded maximum in check (5.78), followed by zinc sulphate (5.39) and *T.viride* (5.16). Among the bio-agents, *P. fluorescens* reduced disease by 25.96% over the check. Among the abiotic agents, salicylic acid recoded superior in all treatments over the control which reduced disease 38.65% over the check followed by calcium sulphate (32.50%) and minimum reduction was observed in zinc sulphate (6.73%) followed by borax (15.96%).

The average percent disease index was observed minimum in Metalaxyl 0.3% was 28.80% having disease reduction 40.88% (Table 4). However, the different concentrations of Metalaxyl were found at par with each other. Maximum average disease index was recorded in check (48.71) followed by *T.viride* (48.36) and *P. fluorescens* (48.09). Among the abiotic agents, salicylic acid reduce the disease by 30.11% over the check followed by calcium sulphate (22.54%) and minimum disease reduction was recorded in zinc sulphate was 5.93%.

Tirmali and Kolte (2012) reported the efficacy of several plant defense activators in the management of *A. candida* pathogen including calcium sulphate, potassium chloride, potassium sulphate, zinc sulphate and borax significantly reduced the pustules size in comparison to control.

Sharma and Kolte (1994) reported that potassium fertilized

plants exhibited 30 to 45 per cent less disease severity of *Alternaria* blight based on the number and size of the spots, average disease index on leaf and pods. Tewari (1991) found that foliar application of the calcium reduce the per cent disease severity of *Alternaria* blight in rapeseed. Antonova *et al.* (1984) and Dixon *et al.* (1987) reported that boron application in the cabbage increase resistance to club root. Singh *et al.* (2020) reported that the size of pustule of white rust was recorded minimum in Metalaxyl 0.3% was 0.32 mm followed by Metalaxyl 0.2% (0.33 mm) in cv. RH-749 during 2015-16 while maximum size of pustules were observed in check was 6.55 mm followed by zinc sulphate at 0.50% (5.88 mm) during 2016-17. Among the abiotic chemicals, salicylic acid was recorded significantly better over all the treatments. The number of pustules were recorded maximum in the check was 7.47 and 7.53 followed by zinc sulphate at 0.50 % was 7.13 and 7.27 during 2015-16 and 2016-17, respectively. Salicylic acid 0.25% reduce 31.50% white rust disease over the control followed by calcium sulphate at 1.00% which reduce disease 23.99%. Zinc sulphate at 0.50% was found least effective abiotic chemical which reduce 6.14% disease.

Conclusion

Among abiotic agents, salicylic acid, calcium sulphate and potassium chloride were found effective in reduction of white rust disease in Indian mustard. While, *P. fluorescens* was found best among the bio agents in reducing the disease over the check. However, the number of pustule was observed minimum in Metalaxyl 0.2% and Metalaxyl 0.3% in comparison to abiotic agents and bio-agents.

References

- Antonova TS, Zaichlor VE and Kalinchenlw TV. 1984. Role of calcium in the resistance of the sunflower anthodium to gray root. *Sciskokhozyaistvernnyaya Biologiya* **11**:67-68.
- Benhamon N and Garand C. 2001. Cytological analysis of defense related mechanisms in pea root tissues in response to colonization by nonpathogenic *Fusarium oxysporum* Fo47. *Phytopathol* **91**: 730-740.
- Biswas C, Singh R and Kumar PV. 2011. Dynamics of white rust disease in mustard (*B.juncea*) in relation to date of sowing and weather parameters. *Indian J Agril Sci* **81**: 1187-1190.
- Dixon GR, Naiki T, Webster A and Wilson F. 1987. Integrated use of boron calcium cyanamide and nitrogen for control of club root (*Plasmodiophora brassicae*), In: *Crop Protec Northern Britan* 399-404.
- Droly S, Vinokur V, Wiess B, Cohen L, Daws A, Goldschmidt EE and Porat R. 2002. Induction of

- resistance to *Pennicilium digitatum* in grape fruit by the yeast bio-control agent *Candida oleophila*. *Phytopathol* **92**: 93-399.
- Durrant WE and Dong X. 2004. Systemic acquired resistance. *Annu Rev Phytopathol* **42**: 185-209.
- Gorlach J, Volrath S, Oostendorp M, Kogel KH, Beckhove U, Staub T, Ward E, Kessmann H and Ryals J. 1996. Benzothiadiazole, a novel class of inducers of systemic acquired resistance, activates systemic resistance in wheat. *Plant Cell* **8**: 629-643.
- Hammerschmidt R. 1999. Induced disease resistance: how do induced plants stop pathogens. *Physiol Mol Plant Pathol* **55**: 77-84.
- Howell CR, Hanson, LE, Spitanovic RD and Puckhaber CS. 2000. Induction of terpenoid synthesis in cotton roots and control of *Rhizoctonia solani* by seed treatments with *T. virens*. *Phytopathol* **90**: 248-252.
- Jones JDG and Dangl JL. 2006. The plant immune system. *Nature* **444**: 323-329.
- Kessmann H, Staub T, Hofmann C, Maetzke T, Herzog J, Ward E, Uknes S and Ryals J. 1994. Induction of systemic acquired disease resistance in plants by chemicals. *Annu Rev Phytopathol* **30**: 439-459.
- Kombrink E and Sommsich IE. 1995. Defence responses of plants to pathogens. *Adv Bot Res* **21**: 1-34.
- Liu L, Kloepper JW and Tuzun S. 1995. Induced systemic resistance in cucumber against bacterial angular spot by plant-growth promoting rhizobacteria. *Phytopathol* **85**: 843-847.
- Matheron ME and Porchas M. 2002. Suppression of Phytophthora root and crown rot on pepper plants treated with Acibenzolar-SMethyl. *Plant Dis* **86**: 292-297.
- Mathur P, Sharma E, Singh SD, Bhatnagar AK, Singh VP and Kapoor R. 2013. Effect of elevated CO₂ on infection of three foliar diseases in oilseed *B. juncea*. *J Plant Pathol* **95**: 135-144.
- Panse VG and Sukhatme PV. 1985. Statistical Methods for Agricultural Workers, 4th ed., ICAR, New Delhi, 347 p.p
- Romero AM, Kousik CC and Ritchie DF. 2001. Resistance to bacterial spot in bell pepper induced by acibenzolar-5-methyl. *Plant Dis* **85**: 189-194.
- Safari S, Soleimani MJ, Mohajer A and Fazlikhani L. 2013. Possible structure-activity profile of salicylate derivatives: their relationship on induction of systemic acquired resistance. *J Agril Technol* **9**: 1215-1225.
- Sharma SR and Kolte SJ. 1994. Effect of soil applied NPK fertilizers on severity of black spot disease (*Alternaria brassicae*) and yield of oil seed rape. *Plant and Soil* **167**: 313-320.
- Singh H, Ratnoo RS, Trivedi A, Jain HK, Saharan V and Sharma FL. 2020. Effect of plant defense activators on white rust of mustard cv. RH-749 under artificial epiphytotic conditions. *J Oilseed Brassica* **11**: 55-61.
- Singh US, Dought KJ, Nashaat NI, Bennet RN and Kolte SJ. 1999. Induction of systemic resistance to *Albugo candida* in *B. juncea* by pre or co-inoculation with an incompatible isolate. *Phytopathol* **89**: 1226-1232.
- Spletzer ME and Enyedi AJ. 1999. Salicylic acid induces resistance to *Alternaria solani* in hydroponically grown tomato. *Phytopathol* **89**: 722-727.
- Sticher L, Mauch MB and Metraux JP. 1997. Systemic acquired resistance. *Annu Rev Phytopathol* **35**: 235-270.
- Surekha CH, Neelapu NNR, Siva Prasad B and Sankar GP. 2014. Induction of defense enzymes and phenolic content by *Trichoderma viride* in *Vigna mungo* infested with *Fusarium oxysporum* and *Alternaria alternata*. *Intl J Agril Sci Res* **4**: 31-40.
- Tewari JP. 1991. Current understanding of resistance to *Alternaria brassicae* in crucifers. *Proc. GCRIC, 8th Intl Rapeseed Congr.* July 9- 11, Saskatoon, Canada, **2**: 471-476.
- Thakur M and Sohal BS. 2013. Role of elicitors in inducing resistance in plants against pathogen infection: A Review. Hinadwi Publishing Corporation, ISRN Biochemistry, 1-10.
- Tirmali AM and Kolte SJ. 2012. Induction of host resistance in mustard with non-conventional chemicals against white rust (*Albugo candida*). *J Pl Dis Sci* **7**: 27-31.
- Verhagen BWM, Glazebrook JT, Chang HS and Van Loon LC. 2004. The transcriptome of Rhizobacteria induced systemic resistance in *Arabidopsis*. *Mol Plant Microbe Interact* **17**: 985-908.
- Vishwanath C, Kolte SJ, Singh MP and Awasthi RP. 1999. Introduction of resistance in mustard (*B. juncea*) against *Alternaria* black spot with an avirulent *Alternaria brassicae* isolate-D. *Euro J Plant Pathol* **105**: 217-220.
- Wheeler BEJ. 1969. An Introduction to Plant Diseases. John Wiley and Sons, London, UK.
- Yalpani N and Raskin I. 1993. Salicylic acid: a systemic signal in induced plant resistance. *Trends Microbiol* **1**: 88-92.