

Development of dwarf *B. carinata* genotype employing inter-specific hybridization between *B. juncea* and *B. carinata* and cyto-morphological studies for patterns of variation

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Abstract

Successful inter-speciûc hybrid plants were obtained through sexual hybridization between *Brassica juncea* cv. NRCDR-2 (2n=4x=36, AABB), and *B. carinata*, NRCKR-304 (2n=4x=34, BBCC) using the latter as a pollen parent. Morphological and cytological analyses were carried out to conûrm the hybrid nature of F_1 plants. The F_1 plants (2n=35) were intermediate for most of the morphological attributes. Although, the F_1 s showed poor pollen fertility, nevertheless, few seeds were obtained from open pollination. Meiotic analysis of F_1 plants showed a predominance of univalents, a typical feature of wide hybrids. The occurrence of chromosome association ranging from bivalents (0–16), trivalent (0–1) and quadrivalent (0–1) in the F_1 s indicated homeologous pairing between the AB and BC genomes. The study suggests that *B. carinata* has partial genome homeology with *B. juncea*. Patterns of variations in F_2 progenies of the cross were estimated. In the F_2 generation, more number of plants resembling *B. juncea* was observed indicating transgressive segregation. Present study also indicated that inter-speciûc hybridization in Brassica could be a potential source for generating variability besides broadening the genetic base and segregants with desirable agro-economical characteristics were selected for further advancement and evaluation.

Key words: cytomorphology, homeologous pairing, interspeciûc hybrids, meiotic analysis, wide hybridization.

Introduction

Brassica is economically one of the most important genus of the family Brassicaceae. It constitutes important sources of vegetables, cooking oil and condiments (Cardoza and Stewart, 2004). Narrow genetic variability in crop Brassicas, caused mainly due to intensive selection over past several decades, has jeopardized the crop improvement programmes (Cowling, 2007; Ananga et al., 2008). Fortunately, inter-speciûc hybridization offers great potential for the improvement of Brassica crop and is being widely utilized for expanding genetic variability, introgressing nuclear genes that conferred desirable agronomic traits or cytoplasmic genes for inducing male sterility, Chromosome addition lines have also been generated to locate genes on speciûc chromosomes and for construction of genetic maps, to elucidate homoeology between different genomes of Brassica (Prakash et al., 2009). Such crosses provides immense opportunities for generating genetic diversity and broadening the genetic base (Choudhary et al., 2002; Choudhary and Joshi, 2012a; 2012b) and are also useful to transfer valuable characters across the species (Sharma and Singh, 1992).

Brassica species are three basic diploid species, specifically B. rapa (AA, 2n=20), B. nigra (BB, 2n=16) and B. oleracea (CC, 2n=18), and three amphidiploids, each of which evolved as a natural allo-tetraploid following hybridization between pairs of the three diploids, namely B. napus (AACC, 2n=38), B. juncea (AABB, 2n=36) and B. carinata (BBCC, 2n=34) (Nagaharu, 1935). B. juncea (L.) Czern & Coss. (Indian mustard) is the predominantly cultivated Brassica species in the Indian subcontinent with yield potential of 15–30 g ha-1 (Meena et al., 2017). Although B. juncea is well adapted to drier conditions and mature earlier than other oilseed Brassica species (Kimber and McGregor, 1995). However, this species is susceptible to aphid, Alternaria blight, white rust and have a limited genetic variation for resistance to both biotic and abiotic stresses (Kumar et al., 1997).

On the other hand, Ethiopian or Abyssinian mustard (*B. carinata* A. Braun, 2n=34) possesses has several agronomical important traits that are rare in other Brassica oil crops such as non-dehiscent siliquae and a much more developed and aggressive root system, heat, drought tolerance, tolerance to various biotic and abiotic stresses and availability of yellow-seeded germplasm (Jiang *et al.*,

2007) which makes potential oilseed crop under semi arid conditions as winter crop. Inspite of these positive attributes, it could not find favour with farmers because this crop suffers from many agronomic limitations like longer crop duration, poor harvest index, low oil content, short shoot length and small pod size *etc*.

Thus, both B. juncea and B. carinata species have desirable characteristics and deûciencies. In view of these facts, the present investigation was conducted with a basic objective to combine the useful genetic attributes of B. juncea and B. carinata, to create genetic variation through inter-speciûc hybridization and possible introgression of genes with useful agro-morphological traits of economic importance. While attempting interspeciûc and inter-generic crosses many have necessitated intervention in the forms of ovary culture, embryo rescue and protoplast fusion (Warwick and Black, 1993; Rieger et al., 1999) to overcome compatibility barriers. Here, we report successful synthesis of interspeciûc hybrids between B. juncea (cv. NRCDR-2) and B. carinata (NRCKR-304) by sexual matings followed by their characterization in terms of morphological attributes, genome homology and differentiation pattern based upon crossability, meiotic behaviour of chromosome and fertility factors of the parents and their F₁s. Additionally, patterns of variations in F2 derivatives and subsequent development of dwarf B. carinata genotype from segregants is also being reported.

Materials and Methods Plant material

Seed samples of *B. juncea* (cv. NRCDR-2) and *B. carinata* (NRCKR-304) were obtained from the germplasm section of the ICAR-Directorate of Rapeseed-Mustard Research, Bharatpur.

Hybridization

Reciprocal inter-specific crosses between plants of the cultivated B. juncea (cv. NRCDR-2) and B. carinata (NRCKR-304) were attempted to produce inter-specific hybrids. Unopened flower buds of B. juncea (cv. NRCDR-2) were emasculated in the afternoon, covered with paper bags and pollinated with freshly collected pollens of B. carinata (NRCKR-304) in the following morning and covered again. The seeds collected from the crossed plants and parents were sown in earthen pots for evaluation and further advancement of progenies from F_2 generation grown field conditions. Morphological comparisons were made for identifying the F_1 s and ascertained cytologically.

Cytological analysis

For meiotic observations flower buds of an appropriate size were collected from mature plant and fixed in freshly prepared carnoy's fluid (ethanol: chloroform: acetic acid - 6:3:1), supplemented with a drop of ferric chloride solution, for a minimum of 24 hours at room temperature and subsequently stored in 70% alcohol at 10°C. For meiotic analysis anthers were squashed in 1% acetocarmine and a total of 30 PMCs were analyzed at diakinesis/metaphase I stages of meiosis in F₁ hybrid plants. For percentage pollen stainability, the pollen grains were stained in 1:1 (glycerine: acetocarmine) mixture and on an average five slides were scored for stainable pollen grains. Normal pollen grains were round, densely stained and were distinguishable from small, shrunken and lightly stained sterile pollen grains.

F, and generation advancement

The seeds harvested from F₁ plants were sown in the field. Based on morphological attributes such as plant appearance in comparison to parent species, leaf shape, size, colour and attachment to the stem, and inflorescence characteristics of the parent species, the F, plants were grouped visually into four broad categories: B. juncea type, B. carinata type, intermediate type and any other type. From F_2 segregating progenies four dwarf type B. carinata plants were selected and from F₂ progenies one dwarf type B. carinata genotype DRMR-C-16-6 was selected (< 100 cm) maintained and advanced up to F₄ generations. Comparison of performance of dwarf type B. carinata genotype (DRMR-C-16-6) for three years was made from 2016-17 to 2018-19. Observations on plant height (cm), days to maturity, primary branches per plant, main shoot length (cm), siliquae length, seeds per siliqua, 1000 seed weight (g), Oil content (%) were made.

Results and Discussion Hybridization and morphological characteristics of F₁ hybrids

Inter-specific hybrid derived from cross NRCDR-2 (*B. juncea*) × *B. carinata* (NRCKR-304) through sexual hybridization was confirmed through cyto-morphological studies. Inter-specific hybrid plants were obtained when *B. carinata* was used as a male parent, and all the hybrids grew up to maturity. NRCKR-304 is a registered germplasm for early maturity, long main shoot and bold seed. One hundred and Thirty eight buds were pollinated from which 20 siliquae with a total of 29 seeds were obtained. Out of the 29 seeds only three the plants, were found to be hybrid and thus the percentage success of crossability was 10.34%. While attempting inter-speciûc/inter-generic

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Characteristics	B. juncea (cv. NRCDR-2)	B. carinata (NRCKR-304)	F ₁ hybrids
Plant height(cm)	195	155	85
Days to 50% flowering	46	38	41
Days to maturity	147.5	132	127
Primary branches per plant	6.8	6.5	7.2
Secondary branches per plan	nt 20.5	17.3	17.8
Main raceme length (cm)	82.6	85.4	60
Siliqua length (cm)	5.6	3.7	2.9
Seeds per siliqua	15.7	16.0	10.2
1000 seed weight (g)	5.3	5.6	2.8
Corolla length (cm)	0.7	1.2	1.4
Corolla width (cm)	0.3	0.7	0.7
Siliqua texture	Smooth	Smooth	Undulated
Seed colour	Dark Brown	Dark Yellow	Light brown
Leaf colour	Medium green	Medium green	Light green
Leaf hairiness	Sparse	sparse	Sparsely hairy
Leaf margin	Serrate	Entire	Dentate
Leaf lobes	Present	Absent	Present

Table 1: Plant morphological characteristics of parents and the F₁ hybrids of B. juncea (cv. NRCDR-2) × B. carinata (NRCKR-304)

crosses most of authors necessitated the intervention in the forms of ovary culture, embryo rescue and protoplast fusion (Warwick and Black, 1993; Rieger et al., 1999) to overcome compatibility barriers.

Yellow

41.541.8

Petal colour

Oil content (%)

The F, hybrid plants (2n=20) obtained were found to be intermediate and vigorous in terms of many phenotypic features and inûorescence attributes of both the parents. These observations are in congruence with earlier reports (Choudhary and Joshi, 2012a, b; Choudhary et al., 2000). The hybrid plants were medium in height, profusely branched and intermediate to their parents for most of the morphological and inflorescence attributes. The leaves were petiolate, lobed and lyrately pinnatified. The dentation of leaf margin was noted to be crenate type with acute tip and petal colour (light yellow) was intermediate to the parents. The seeds of hybrids were small in size and light brown in colour. The hybrid plants had smaller siliquae as compared to the parents (Table 1). The present investigation reports the successful hybridization between B. juncea and B. carinata even when it is involved as a male parent. It is quite useful in maintaining the cytological background of the crop species in distant hybridization programmes. The occurrence of characteristics from both progenitor species in the hybrids indicates that the F, plants inherited genomes of both parental species crossed. This is an advantage, since it would allow better selection for speciûc attributes in segregating progenies.

Cytology of F₁ hybrids

Yellow

40.03

Meiotic data of F, hybrids has been summarized in Figure 1, Table 2 and Table 3. Cytological analysis of F, hybrids (2n = 35) not only confirmed their hybridity but also indicated extent of genome homoeology between the parents. Meiotic analysis of the F, hybrids of B. juncea and B. carinata showed a mixture of univalents, bivalent, trivalent and quadrivalent in a total of 30 PMCs analyzed. The F₁ hybrids showed predominance of univalents, which is a typical feature of wide hybrids (Kumar et al., 2013, 2015, 2018). The bivalents ranged between 02 to 16 in various PMCs. The average chromosome association in the hybrid was of 0.13IV + 0.19III + 7.69II + 18.63I. The number of univalents and bivalents ranged from 3 - 27 and 2 - 16, respectively, whereas number of trivalent and quadrivalent never exceeded one per PMC. Chromosome pairing in the F, hybrids could be interpreted as a result of auto as well as allosyndesis within and between, respectively, the AB and BC genomes (Choudhary and Joshi, 2012a, Prakash, 1974). From the observations the occurrence of chiasmatic and heteromorphic pairing, multivalent associations in the form of trivalents and quadrivalents, though only in a few PMCs, and more importantly, the formation of up to 16 bivalents could be accounted for by autosyndesis within the AB and BC genomes, clearly indicating homoeologous pairing between chromosomes of AB and BC genomes. Thus, it appears B. carinata has some homoeology with B. juncea

Light yellow

41.3

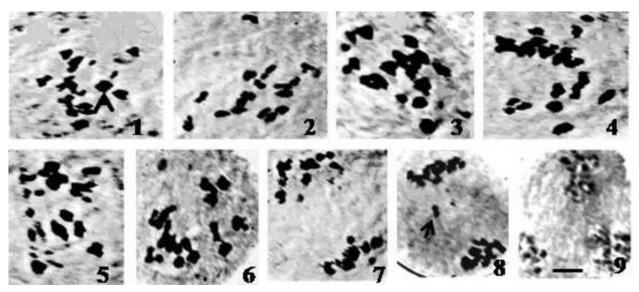


Fig. 1. Meiotic analysis of F₁ hybrids showing chromosome associations at diakinesis/metaphase I and chromosome distribution at anaphase I. 1. Metaphase I 1IV + 12II + 7I (Quadrivalent marked by arrow head), 2. Metaphase I 6II + 23I, 3. Diakinesis 10II + 15I, 4. Diakinesis 7II + 21I, 5. Metaphase I 8II + 19I, 6. Diakinesis 6II + 23I, 7. Anaphase I with laggards (marked by arrow), 9 Anaphase II (showing intermixing of chromatids at one pole), Bar: 10 μm.

Table 2: Chromosome configuration at diakinesis/metaphase I in PMCs of F_1 hybrids B. juncea (cv. NRCDR-2) × B. carinata (NRCKR-304) hybrids (2n=35)

	Chromosome configuration (number/PMC)						Number of PMCs observed		
_	IV	I	II	II		I		Number	Frequency (%)
	1		_	6		19		1	3.3
	1		_	12		7		1	3.3
			1	3		26		1	3.3
			1	6		21		1	3.3
			1	4		24		1	3.3
				16		3		1	3.3
				13		9		1	3.3
				12		11		1	3.3
				10		15		1	3.3
				9		17		3	10.0
				8		19		1	3.3
				7		21		2	6.7
				6		23		8	26.7
				5		25		2	6.7
				4		27		3	10.0
				2		31		2	6.7
Total								30	
Range	0-1	0	-1	2-16		3-27			
Mean ± SEm	0.7	0.04	0.1	0.05	7.7	3.9	18.6	7.6	

indicating possibility of gene inrogression through conventional means. Numerous disjunctional abnormalities including late disjunction of bivalents, and laggards were observed at anaphase I and II (Table 2) was recorded in a few cells, which is similar to the study reported by Choudhary and Joshi (2012b). Majority of PMCs (74%) analyzed showed laggards univalents or bivalents at anaphase I and II, where as only 25.9%

Table 3: Chromosome distribution at anaphase I and II in hybrids of B. juncea (cv. NRCDR-2) × B. carinata (NRCKR-304) hybrids (2n=35)

Chromosome distribution	PMCs observed	
	Number	Percentage
Equal distribution	7	25.93
Laggards at A I	15	55.56
Laggards at A II	5	18.51
Total	27	

showed equal distribution of chromosomes at anaphase I. However, a few cells were recorded with normal distribution of chromosomes resulting in some fertile pollen grains in the hybrids. The drastic decrease in pollen fertility (22.4%) and reduced seed set recorded in the present hybrids might be due to meiotic irregularities and segregational anomalies (Singh, 1993). The occurrence of homoeologous pairing between B. juncea and B. carinata chromosomes and few seeds set by interspeciûc hybrids offers an opportunity for the transfer of useful genes across the species.

F, generation

From the ûeld grown F, segregating progenies, a total of 102 plants in which 44 (B. juncea type), 30 (B. carinata), 17(intermediate type) and 11(other type) obtained were used to record observations on different agromorphological traits. Wide phenotypic variations regarding plant types and metric traits were observed in F, populations. The pattern of segregation for plant types showed high recovery of B. juncea type plants (43.31%) as compared to B. carinata type (29.4%), intermediate category (16.6%) and other types (10.9%) (Table 4). High frequency of the maternal and intermediate types of plants in F, generation of B. juncea \times B. rapa crosses was also by Choudhary et al. (2002). In all, five plants were found to be completely male sterile. With regards to pollen fertility of F, progenies recorded higher mean pollen fertility (46.8%) compared F₁ progenies (22.4%).

The occurrence of B. juncea (the maternal parent) type plants in larger frequency compared with other three types in F, generation might resulted from a greater viability of female gametes with higher chromosome number (Subudhi and Raut, 1994). High frequency of the maternal and intermediate types of plants in F₂ generation of B. juncea × B. rapa crosses was also by Choudhary et al. (2002) and Chuodhary and Joshi (2012b). Wide phenotypic variation and transgressive segregation for plant height, Secondary branches per plant, main raceme length etc. in the F₂ generation might have resulted from recombination and or eventual segregation of aneuploid forms arising in the populations. This suggested the possibility of improving the derivatives through selection for these traits. This could be explained why Olsson (1960a, 1960b) found wide variation in F₂ and F₃ plants of the cross B. rapa \times B. nigra for plant height, ûowering and maturity, siliqua size and pollen fertility. High-yielding physiological variants from segregants of the B. campestris × B. nigra cross are reported by Prakash (1973). On the similar hypothesis, Rao et al. (1993) and Choudhary et al. (2002) reported transgressive segregants for yield components from the F, generation of B. napus \times B. carinata and B. juncea \times B. rapa inte-rspecific crosses.

Generation advancement

From F₂ progenies (dwarf B. carinata type) one dwarf type B. carinata genotype DRMR-C-16-6 was selected (< 100 cm) maintained through selfing and advanced up to F₂ generations. Comparison of performance of dwarf type B. carinata genotype (DRMR-C-16-6) for three years was made from 2016-17 to 2018-19. Observations on plant height (cm), days to maturity, primary branches per plant, main shoot length (cm), siliquae length, seeds per siliqua, 1000 seed weight (g), Oil content (%) were made (Table 5) along with B. carinata (cv. Kiran), and both the parents NRCKR-304 (B. carinata) and B. juncea (cv. NRCDR-2) respectively. Mean values of plant height (83.7 cm), days to maturity (128.7), Secondary branches (22.1) and oil content 41.4% were found to be higher when compared with parents B. carinata (NRCKR-304) and B. juncea (cv. NRCDR-2). Dwarf genotype B. carinata (DRMR-C-16-6) was observed to be superior almost all agromorphological characteristics when compared with the variety of B. carinata (Kiran) (Table 5). Thus present

Table 4: Segregation for plant types in the F, generation of B. juncea (cv. NRCDR-2) and B. carinata (NRCKR-304) cross

Cross	Plant type frequency				
	B. juncea (cv. NRCDR-2)	B. carinata (NRCKR-304)	Intermediate	Other	Number of Plants observed
B. juncea (cv.NRCDR-2) ×B. carinata (NRCKR-304)	44(43.1)*	30(29.4.3)	17(16.6)	11(10.9)	102

^{*}Numbers in parenthesis indicate percentage

Characteristics	DRMR-C-16-6 (B. carinata)	NRCKR-304* (B. carinata)	Kiran (B. carinata)	NRCDR-2** (B. juncea)
Days to maturity	128.67	136	162.67	143.53
Plant height (cm)	83.66	158.33	234	206
Primary branches	10.3	5.27	9.17	6.47
Secondary branches	22.13	13.7	21.3	15.53
Main shoot length (cm)	58.33	89.5	38.33	89.1
Siliqua length (cm)	3.83	3.7	3.4	5.27
Seeds per siliqua	15.43	16.3	12.7	15.97
1000 seed weight (g)	4.77	5.17	3.23	5.25
Oil content (%)	41.43	40.44	39.0	41.4

Table 5: Comparison of performance of dwarf type *B. carinata* genotype (DRMR-C-16-6) for three years from 2016-17 to 2018-19 (average data) along with parents

parents = *NRCKR-304, **NRCDR-2

study indicated that inter-speciûc hybridization in Brassica could be a valuable source for generating genetic diversity for broadening base and segregants with desirable attributes can be valuable source for further crop improvement of *B. carinata* and as well as *B. juncea*.

Conclusion

Successful synthesis of interspeciûc hybrids between B. juncea (cv. NRCDR-2) and B. carinata (NRCKR-304) by sexual mating followed by their characterization in terms of morphological attributes, genome homology and differentiation pattern based upon crossability, meiotic behaviour of chromosome and fertility factors of the parents and their F₁s. The study suggests that B. carinata has partial genome homeology with B. juncea. In the F, generation, more number of plants resembling B. juncea was observed indicating transgressive segregation. Present study also indicated that inter-speciûc hybridization in Brassica could be a potential source for generating variability besides broadening the genetic base and segregants with desirable agro-economical characteristics were selected for further advancement and evaluation and subsequent selection of dwarf type B. carinata genotype.

References

- Ananga A, Cebert E, Soliman K, Kantety R, Konan K and Oshieng JW. 2008. Phylogenetic relationships within and among Brassica species from RAPD loci associated with blackleg resistance. *Afr J Biotech* 7: 1287-1293.
- Cardoza V and Neal CS Jr. 2004. Brassica Biotechnology: Progress in cellular and molecular biology. *In vitro Cell Dev Pl* **40**: 542-551.
- Choudhary BR and Joshi P. 2012b. Cytomorphology of *Brassica napus* × *B. rapa* hybrids and patterns of variation in the F, derivatives. *Caryologia* **65**: 316-321.

- Choudhary BR and Joshi P. 2012a. Crossibility of *B. carinata* and *B. tournefortii*, and cytomorphology of their F₁ hybrid. *Cytologia* **77:** 453-458.
- Choudhary BR, Joshi P and Ramarao S. 2002. Cytogenetics of *B. juncea* × *B. rapa* hybrids and pattern of variation in the hybrid derivatives. *Plant Breed* **121**: 292–296.
- Choudhary BR, Joshi P and Singh K. 2000b. Synthesis, morphology and cytogenetics of *Raphano fortii* (TTRR, 2n=38): a new amphidiploid of hybrid *B. tournefortii* (TT, 2n=20) × *Raphanus caudatus* (RR, 2n=18). *Theor Appl Genet* **10:** 990-999.
- Cowling WA. 2007. Genetic diversity in Australian canola and implications for crop breeding for changing future environments. *Field Crop Res* **104**: 103-111.
- Jiang Y, Tian, Li, Chen, L and Meng J. 2007. Genetic diversity of *B. carinata* with emphasis on the interspeciûc crossability with *B. rapa. Plant Breed* **126**: 487–491.
- Kimber DS and McGregor DI. 1995. The species and their origin, cultivation and world production. In: *Brassica oilseeds, production and utilization*. Ed. Kimber D, McGregor DI and Wallingford. CAB International. 1–7.
- Kumar A, Singh BK, Meena HS, Sing, VV, Singh YP and Singh D. 2015b. Cytomorphological and molecular characterization of F₁ hybrids between *B. tournefortii* and *B. rapa*. Cytologia **80**: 317-326.
- Kumar A, Meena HS, Ram B, Priyamedha, Sharma A, Yadav S, Singh VV and Rai PK. 2018. Some cytomorphological evidence for synthesis of inters-peciûc hybrids between *B. juncea* and Autotetraploid *B. fruticulosa*. *Cytologia* **83**: 421–426.
- Kumar A, Singh BK, Singh VV and Chauhan JS. 2013. Cytomorphological and molecular evidences of

- synthesis of interspecific hybrids between *Brassica* rapa and *B. fruticulosa* through hybridization. *Aus J crop sci* **7:** 849-854.
- Kumar PR, Singh D and Chandra N. 1997. Advances in rapeseed mustard breeding. In: *Plant breeding and crop improvement* Vol I. (eds.) Kapoor RL and Saini ML, New Delhi: CBS Publishers and Distributors, 104–129.
- Meena HS, Kumar A, Singh VV, Meena PD, Ram B and Kulshrestha S. 2017. Genetic variability and interrelation of seed yield with contributing traits in Indian mustard (*B. juncea*). *J Oilseed Brassica* 8: 131–137.
- Olsson G. 1960a. Species crosses within the genus Brassica. I. Artiûcial *B. juncea. Hereditas* **46**: 171–223.
- Olsson G. 1960b. Species crosses within the genus Brassica. II. Artiûcial *Brassica napus* L. *Hereditas* **46**: 351–386.
- Prakash S. 1974. Haploid meiosis and origin of *B. tournefortii* Gouan. *Euphytica* **23**: 591–595.
- Prakash S, Bhat SR, Qurios CF, Kirit PB and Chopra VL. 2009. Brassica and its close allies: cytogenetics and evolution. *Plant Breed Rev* **31**: 21-187.
- Prakash S. 1973. Non-homologous meiotic pairing in the A and B genomes of Brassica: its breeding significance in the production of variable amphidiploids. *Genet Res* 21: 133–137.
- Rao MVB, Babu VR, Radhika K. 1993. Introgression of earliness in *B. napus* L. I. An interspeciûc *B. juncea* and *B. napus* cross. *Intern J Trop Agri* 11: 14–19.

- Riege MA, Preston C and Powle SB. 1999. Risks of gene flow from transgenic herbicide-resistant canola (*B. napus*) to weedy relatives in southern Australian cropping system. *Aust J Agric Res* **50**: 115-128.
- Sharma TR and Singh BM. 1992. Transfer of resistance to *Alternaria brassicae* in *B. juncea* through interspeciûc hybridization among Brassicas. *J Genet Breed* **46**: 373–378.
- Singh RJ. 1993. Plant Cytogenetics. CRC Press. Inc. Bocca Raton.
- Subudhi PK and Raut RN. 1994. White rust resistance and its association with parental species type and leaf waxiness in [B. juncea (L.) Czern & Coss.] and B. napus L. crosses under the action of EDTA and gamma-rays. Euphytica 74: 1–7.
- Nagaharu U. 1935. Genome analysis in Brassica with special reference to the experimental formation of *B. napus* and peculiar mode of fertilization. *Japan J Bot* **7**: 389–452.
- Warwick SI and Black LD. 1993. Guide to the wild germplasm of Brassica and allied crops, Part III: Interspeciûc and intergeneric hybridization in the tribe *Brassiceae* (Cruciferae). Technical Bulletin 1993-16E. Centre for Land and Biological Resources Research, Agriculture and Agri-Food Canada, Ottawa.